

Main Steps in Bacteriological Diagnosis

The diagnostic cycle

Laboratory testing – series of events:

- I. Pre-analytical
- II. Analytical = observation & isolation & identification & enumeration
- III. Post-analytical

Performance must be monitored throughout the entire cycle for quality assurance i.e. accurate, reliable results

Clinical & bacteriological diagnosis of infectious diseases

PRE-ANALYTICAL:

Patient consulted by physician ▼

Physician → tentative clinical diagnosis ▼

Specimens: Collection & labels on containers ▼

Request form + specimens ▼

Lab receives samples; data recorded

ANALYTICAL ► ►

Direct examination: smears, stains ►

POST-ANALYTICAL: Final report elaborated & sent to physician

Subcultures + results of identification systems ▲

Cultures examined; identification systems ▲

Culture media selected, inoculated, incubated ▲

Presumptive reports, preliminary results ▲

Pre-analytic phase: Specimen collection

- Crucial for confirming a certain microorganism as cause of the clinically suspected infectious disease
- Improper specimen collection may cause:
 - Failure to recover the microorganism (no growth on culture medium)
 - Incorrect / harmful therapy e.g directed against a comensal / contaminant microorganism

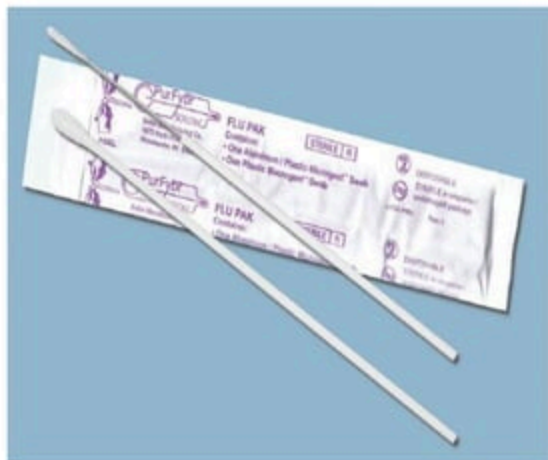
E.g. *Klebsiella pneumoniae*:

- recovered from sputum of pneumonia patient;
- recognised causative agent of pneumonia BUT also may colonize the naso-pharynx
- If sputum sample consisted mostly of saliva then isolating *K.pneumoniae* might not reflect the true cause of the patient's pneumonia but saliva contamination of the sputum sample

Pre-analytic phase: Specimen collection (continued)

Rules for correct specimen collection:

1. **Source:** actual infection site; minimal contamination from adjacent tissues, organs, secretions e.g. throat swabs from peritonsillar fossae and posterior pharyngeal wall, avoiding contact with other oral areas
2. **Optimal moment:** depending on the natural history and pathophysiology of the infectious process e.g. Typhoid fever: blood – 1st week; feces and urine – 2nd-3rd week
3. **Sufficient quantity**



Pre-analytic phase: Specimen collection (continued)

Rules for correct specimen collection (continued):

4. Appropriate collection devices, containers

+ transport systems (container ± transport medium): main objective to decrease time between collection and inoculation to prevent lack of recovery of certain bacteria

5. Sample collection before antibiotics (if possible)

6. Smears performed to supplement culture (if possible)

- Assessment of inflammatory nature of specimen → aid the clinical integration (meaningfulness) of the culture result
- Gram smears e.g. Gram negative bacilli + no growth on aerobic culture (wrong atmosphere or wrong media i.e. fastidious microbes e.g. *Legionella*)

Pre-analytic phase: Specimen collection (continued)

Rules for correct specimen collection (continued):

7. Labeling of specimen containers & **Request form**:

- Legible
- Minimum information:
 - Patient name; identification number (hospital file, practice log book, etc)
 - Source of specimen; clinician + contact data (phone no)
 - Date and hour of collection
 - Clinical diagnosis (suspected infection)
 - Treatments (antibiotics?...)

Pre-analytic phase: Transport

Main transport related objectives:

- Sample related: Transport media
 - Maintain the sample as similar to its original state as possible
- Human & environment related: Packaging and transport systems and regulations
 - Prevent contamination/infection of healthcare staff & environment & general population (biosafety)

Pre-analytic phase: Transport (continued)

Transport media



Pre-analytic phase: Transport (continued)

"Triple" Packaging of biological samples:

- Outer box (usually cardboard, rigid, secure closure system, adequately labeled to state content)
- Inner container (waterproof, resistant to pressure, usually plastic, securely closed by lid, contains additional materials to absorb shocks e.g. bubbled plastic bags and leakages e.g. absorbent material)
- Sample containers (tubes, plates) inserted in the inner plastic container, wrapped in above mentioned shock- and fluid absorbent materials
- + request form and other documents inserted in sealed plastic bags, inserted in outer box or stucked to its exterior

Triple packaging

1

Primary receptacle
Leakproof



+

2

Secondary packaging
resistant 95 kPa

Liquid absorbent



+

3

Outer packaging



Pre-analytic phase: Transport (continued)

Shipment of biological specimens:
Packaging and transport systems and regulations: IATA (ICAO), ADR...



INSURANCE - If carrier offers insurance, and such insurance is requested in accordance with the conditions thereof, indicate amount insured in figures in box marked "Amount of Insurance".	
SCI	
Nature and Quantity of Goods (incl. Dimensions of Volume)	
Biological Substance, Category B UN337/3 2 x 100ml Packing Instruction 650 Dry ice Class 9 UN1845 1 x 5kg	



Clinical & bacteriological diagnosis of infectious diseases

PRE-ANALYTICAL:

Patient consulted by physician ▼

Physician → tentative clinical diagnosis ▼

Specimens: Collection & labels on containers ▼

Request form + specimens ▼

Lab receives samples; data recorded

ANALYTICAL ► ►

Direct examination: smears, stains ►

POST-ANALYTICAL: Final report elaborated & sent to physician

Subcultures + results of identification systems ▲

Cultures examined; identification systems ▲

Culture media selected, inoculated, incubated ▲

Presumptive reports, preliminary results ▲

Specimen receipt & preliminary observations (continued)

Examples of acceptance/rejection criteria (checklist):

1. Request form & labels contain all info required (check for consistency !!!)
2. Improperly packaged, leaking / broken containers
3. Time from collection to receipt – too long to allow recovery
4. Improper / lack of transport media
5. Insufficient quantity e.g. single swab for multiple requests
6. Overgrown / dried out culture plates
7.etc.....

Each lab must have such a list and share it with collaborators!

Rejecting samples must be avoided as much as possible!

Collection & transport requirements must be shared with clinicians!!!

Specimen receipt & preliminary observations

- Specially designed area / room for receiving and recording samples
- Rules for manipulating samples and accompanying documents (UNIVERSAL PRECAUTIONS):
 - Samples: biological safety cabinet (BSC), personal protective equipment (PPE): lab coat, gloves, eye&respiratory protection
 - Documents – handled by different person / at different stage e.g. either before or after preliminary examination/processing of sample (after removal of gloves & hand washing) – purpose: avoid cross contamination of objects (log record book, computer, pens, etc)

Specimen receipt & preliminary observations (continued)

Preliminary actions upon receipt of specimens

1. Data entry into lab log book/computer database
2. (Unpacking and) visual examination – check for acceptance criteria (see next slide)
+
3. Microscopic examination of direct mounts/stained smears → presumptive diagnosis
4. Sample(s) taken to area/room where the analytical phase begins

Clinical & bacteriological diagnosis of infectious diseases

PRE-ANALYTICAL:

Patient consulted by physician ▼

Physician → tentative clinical diagnosis ▼

Specimens: Collection & labels on containers ▼

Request form + specimens ▼

Lab receives samples; data recorded

ANALYTICAL ► ►

Direct examination:
smears, stains ►

POST-ANALYTICAL: Final report elaborated & sent to physician

Subcultures + results of identification systems ▲

Cultures examined; identification systems ▲

Culture media selected, inoculated, incubated ▲

Presumptive reports, preliminary results ▲

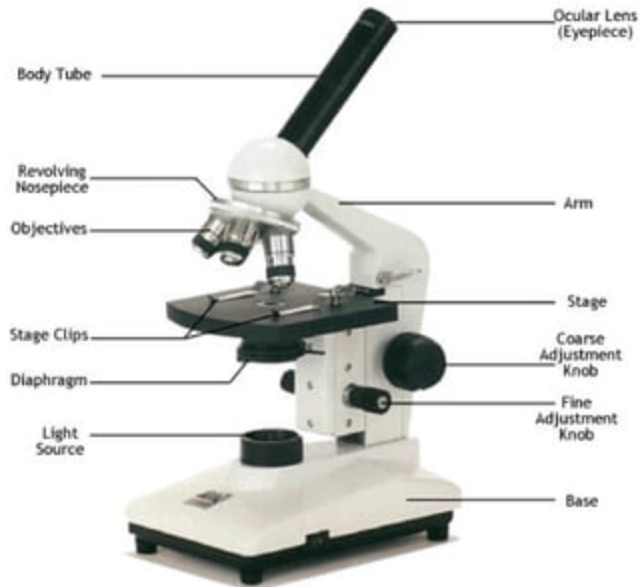
Bacterial infections:
direct identification & characterization methods

- Microscopy
- Cultivation
- Antimicrobial sensitivity

Microscopy

- Types of microscopes
 - Optical - Magnification objectives
 - 10x; 40x; **100x for bacteria**
 - Phase contrast
 - Dark field (dark ground)
 - Fluorescence – UV light
 - Electron

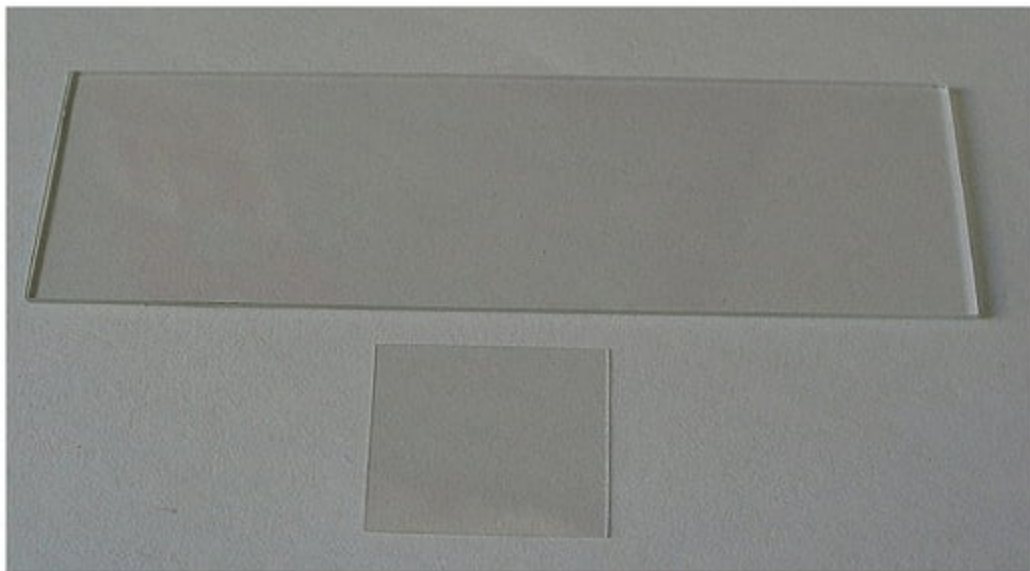
Optical microscope



Microscopic examination

- Wet mounts (unstained materials)
 - Direct light
 - Observation of cells (PMN, macrophages), mobile germs in liquid samples (urine, CSF), shape and disposition of germs (cocci/bacilli/spirilli/vibrios)
- Stained smears

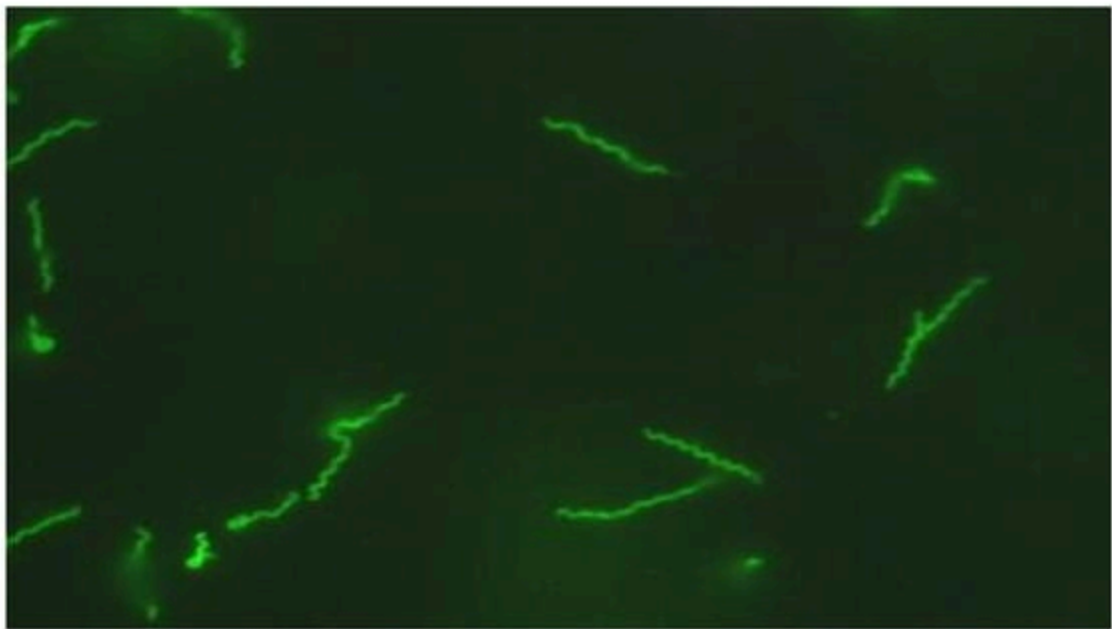
Microscope glass slide and cover slip



Spirochetes – wet mount by dark field microscopy



Treponema denticola – dark field
microscopy + fluorescent dye staining



Stained smears

Main steps:

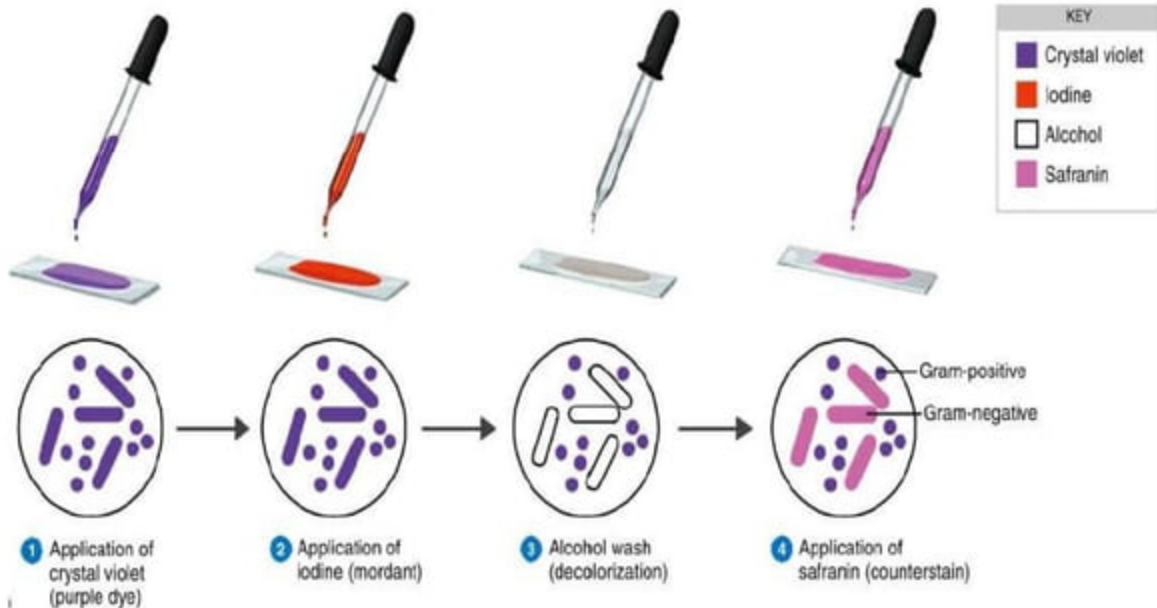
- Smear specimen on microscope glass slide
- Air Drying
- Heat Fixation (flame): help adhesion of specimen to slide, kill bacteria, favour absorption of stain on bacterial surface
- Staining:
 - Monostaining e.g. Methyl blue
 - Combined e.g. Gram, Ziehl Nielsen

Gram staining

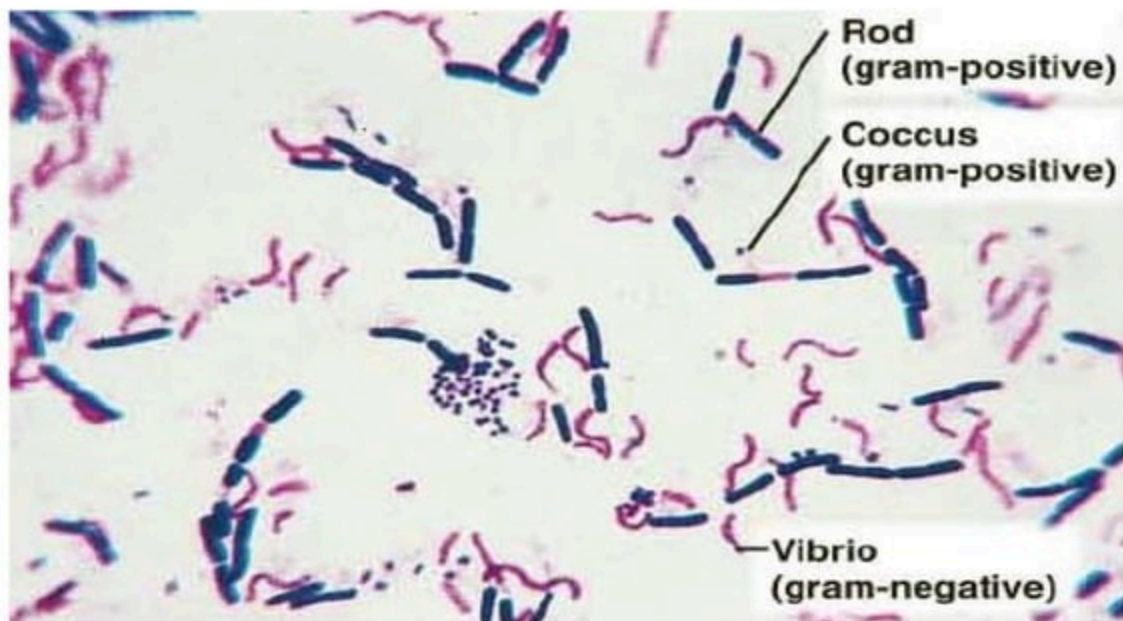
1. heat-fixed smear flooded with crystal violet (**primary stain**)
2. crystal violet is drained off and washed with distilled water
3. smear covered with "Gram's iodine" (Lugol) (**amordant or helper**)
4. iodine washed off: all bacteria appear dark violet or purple
5. slide washed with alcohol (95% ethanol) or an alcohol-acetone solution (**decolorizing agent**)
6. alcohol rinsed off with distilled water
7. slide stained with safranin, a basic red dye (**counter stain**) 2-3 minutes
8. smear washed again, heat dried and examined microscopically

Exact protocol – depending on the kit

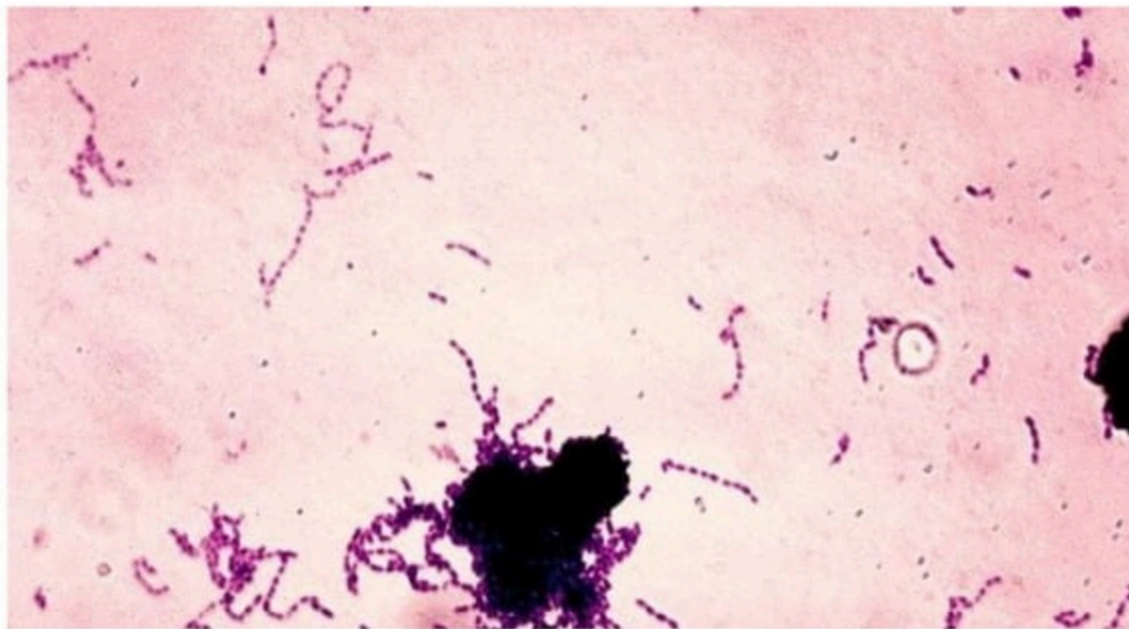
Gram staining



Gram stained smear



Streptococcus mutans – Gram stained smear



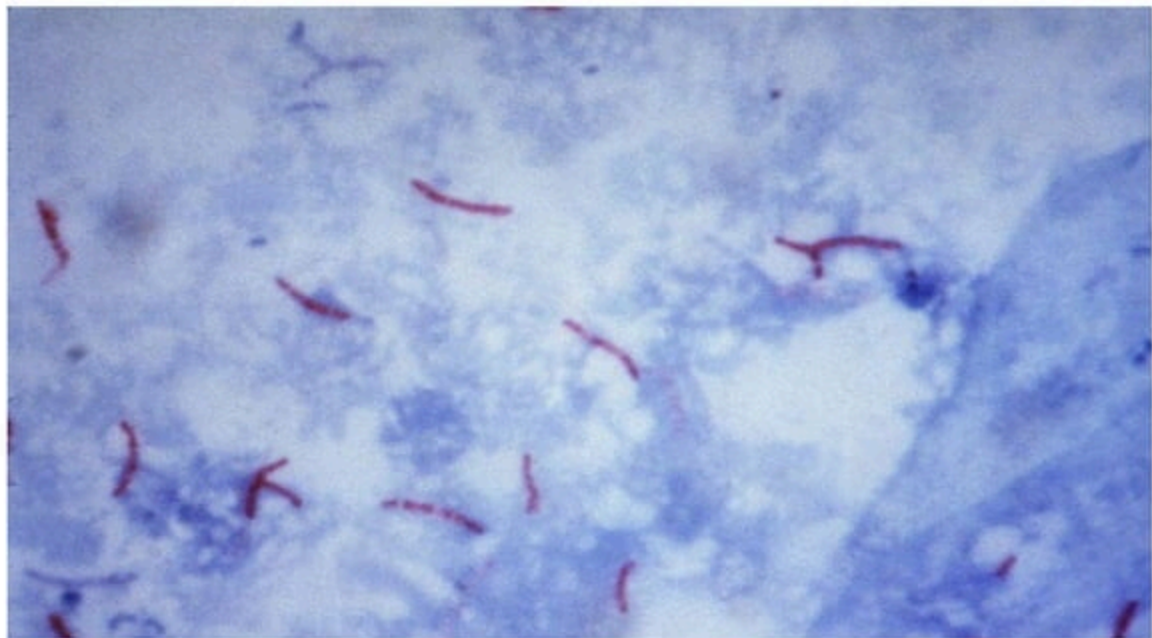
Ziehl-Neelsen Staining

- used to stain *Mycobacterium tuberculosis* and *Mycobacterium leprae* = acid fast bacilli: stain with carbol fuschin (red dye) and retain the dye when treated with acid (due to lipids i.e. mycolic acid in cell wall)

Reagents

- **Carbol fuchsin** (basic dye) - red
- Mordant (heat)
- 20% sulphuric acid (decolorizer) – acid fast bacilli retain the basic (red) dye
- **Methylene blue** (counter stain) – the other elements of the smear, including the background will be blue

Mycobacterium tuberculosis - Ziehl-Neelsen
Staining



Giemsa staining

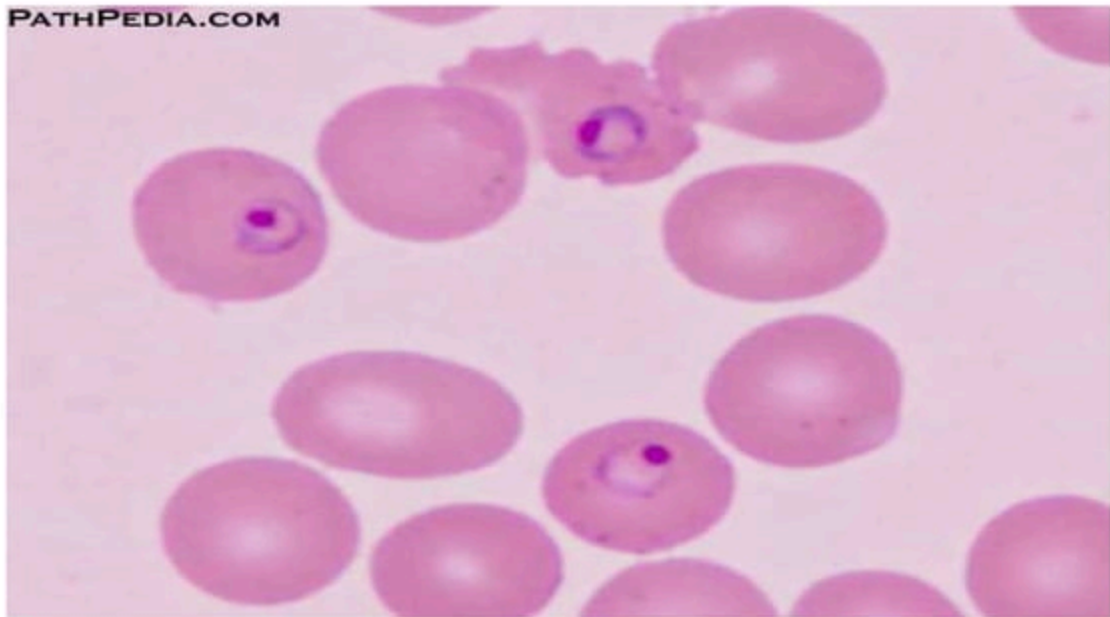
- Smears from blood, vaginal / urethral secretion, bone marrow aspirate

Steps:

- Fixation with methanol (2-3 min)
- Coloration with Giemsa solution
- Washing – buffered water
- Drying
- Microscopic examination

Malaria parasites in blood smear (Wright/Giemsa staining)

PATHPEDIA.COM



Microscopy for various biological specimens

- CSF:
 - wet mounts – assess type & no of cells (white/red blood cells)
 - Stained smears from centrifugation sediment: Gram, Ziehl-Neelsen + additional smear
 - Presumptive causative agents:
 - High no of PMN on wet mount → bacterial meningitis *Neisseria meningitidis*, *Haemophilus influenzae*
 - Ziehl-Neelsen stained smear – very important in case M.tuberculosis is suspected (cultures take 2-3 weeks)

Microscopy for various biological specimens

- Pus
 - Gram stained smears: PMN + *staphylococci*, *streptococci*
- Urine
 - Gram and Ziehl-Neelsen stained smears prepared from sediment (after centrifugation of specimen)
 - Urinary infection: smear with germs + high no of PMN
- Sputum
 - Prewashing of specimen in several, successive Petri dishes (to remove germs from the pharynx attached to sputum)
 - Gram (*staphylococci*, *streptococci*), Ziehl-Neelsen (*M.tuberculosis*)

Bacterial infections:
direct identification & characterization methods

- Microscopy
- **Cultivation** (see presentation on culture media)
- Antimicrobial sensitivity

Cultivation of bacteria on culture media

- Purpose: isolated colonies (single microbial species)
- Identification:
 - Gram stained smears (from colonies)
 - Morphology of colonies (shape, dimensions, margins, colour, ...)
 - Changes in the culture medium (e.g. Hemolysis)
 - Biochemical characters:
 - Enzyme secretion (coagulase, catalase, oxydase)
 - Fermentation of sugars
 - Production of H₂S
 - Immune assays: agglutination, immune fluorescence, ELISA

Bacterial infections:
direct identification & characterization methods

- Microscopy
- Cultivation
- **Antimicrobial sensitivity**

Antimicrobial sensitivity

- ***In vitro*** testing for the sensitivity of microbes to various antibiotics; expressed as:
 - MIC (minimal inhibitory concentration) – the lowest quantity of antibiotic completely inhibiting the multiplication of a bacterial strain
 - MBC (minimal bactericidal concentration) – the lowest quantity of antibiotic able to kill 99.9-100% of the germs of a tested bacterial strain
- ***In vivo*** – concentration of antibiotic at the infection site